

Small molecule biaryl FSH receptor agonists. Part 2: Lead optimization via parallel synthesis

Tao Guo,^{a,*} Anton E. P. Adang,^b Guizhen Dong,^a Dan Fitzpatrick,^a Peng Geng,^a
Koc-Kan Ho,^a Charles H. Jibilian,^a Steven G. Kultgen,^a Ruiyan Liu,^a
Edward McDonald,^a Kurt W. Saionz,^a Kenneth J. Valenzano,^a
Nicole C. R. van Straten,^b Dan Xie^a and Maria L. Webb^a

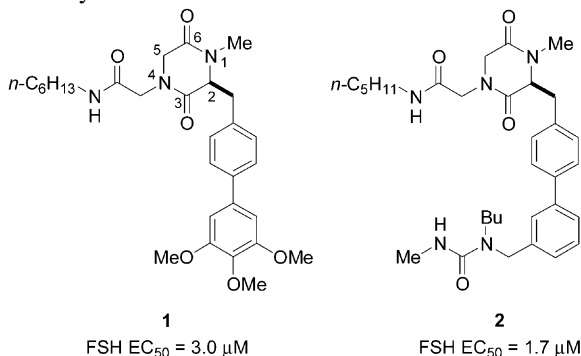
^aPharmacopeia, Inc., PO Box 5350, Princeton, NJ 08543-5350, USA

^bLead Discovery Unit, N.V. Organon, PO Box 20, 5340 BH Oss, The Netherlands

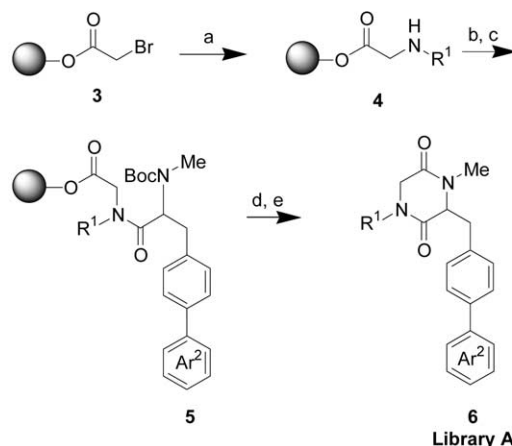
Received 1 December 2003; revised 16 January 2004; accepted 19 January 2004

Abstract—Potent small molecule biaryl diketopiperazine FSH receptor agonists such as **10c** (EC_{50} = 13 nM) and **11f** (EC_{50} = 1.2 nM) were discovered through the design, synthesis and evaluation of three biaryl diketopiperazine optimization libraries with over 300 compounds. These libraries were prepared via solid-phase parallel synthesis using a cyclization-release method.
© 2004 Elsevier Ltd. All rights reserved.

As described in the preceding paper, novel biaryl FSH receptor agonists such as **1** (EC_{50} = 3 μ M) and **2** (EC_{50} = 1.7 μ M) along with a unique combinatorial SAR were discovered through encoded combinatorial synthesis.¹ The results demonstrated that combinatorial synthesis is a powerful tool to aid in the discovery of small molecule agonists for peptide/protein receptors. These biaryl compounds could serve as good starting points in a follow-up optimization program. In this paper, we report the elaboration of **1** and **2** into potent FSH receptor agonists such as **10c** (EC_{50} = 13 nM, MW = 511) and **11f** (EC_{50} = 1.2 nM, MW = 563) via parallel synthesis.



Compounds **1** and **2** share a common biaryl diketopiperazine core, thus the optimization efforts were centered on exploring various side chain substituents on the common core. Specifically, three parallel libraries—A (generic structure **6** in Scheme 1), B (generic structure **10** in Scheme 2), and C (generic structure **11** in Scheme 2)—comprising over 300 compounds were designed to investigate the effect of replacing the metabolically vulnerable side chain amide in **1** and **2** with various



Scheme 1. Synthesis of parallel library A: (a) R^1NH_2 , DMSO; (b) Boc-L or D-N-Me-4-iodophenylalanine, HATU, DIEA, DMF; (c) $Ar^2B(OH)_2$, $Pd(PPh_3)_4$, K_2CO_3 , DMF, 65 °C; (d) TFA, CH_2Cl_2 ; (e) Et_3N , CH_2Cl_2 .

Keywords: FSH Receptor agonists; Parallel synthesis.

*Corresponding author. Tel.: +1-609-452-3746; fax: +1-732-422-0156; e-mail: tguo@pharmacop.com

des-amide fragments (R^1) in combination with introducing a C-5 substituent (R^3) on the diketopiperazine ring as well as varying the aryl substituents (Ar^2 , R^4 , and R^5). The *N*-1-Me group on the diketopiperazine ring, however, was kept unchanged in these libraries because initial analogues of **1** containing other *N*-1-substituents—H, Et, or Pr, for example—all displayed reduced potency ($EC_{50} > 20 \mu M$). In order to prepare the three des-amide libraries, given that the side chain amide nitrogen in **1** and **2** was the resin attachment point in the solid-phase synthesis of the original combinatorial library,¹ a new synthetic route was sought. Eventually, a solid-phase cyclization-release method similar to that reported by Chiron² and Affymax³ researchers was developed (Schemes 1 and 2) to generate these des-amide biaryl diketopiperazine libraries.

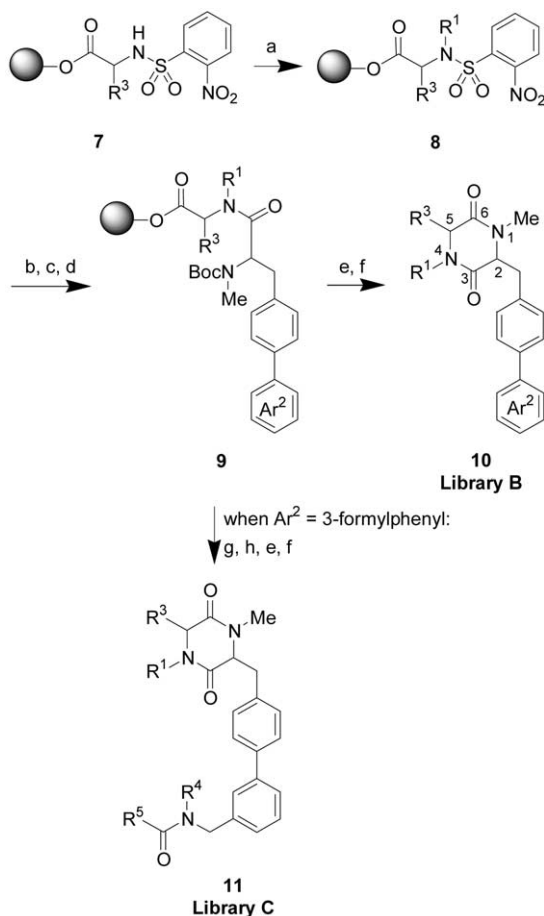
Thus, as shown in Scheme 1, the synthesis of parallel library A began with bromoacetate **3** that was obtained from esterification of TentaGelTM-S-OH resin with bromoacetic acid using 1,3-diisopropylcarbodiimide. Amination of **3** with diverse primary amines R^1NH_2 generated **4**. Acylation of **4** with Boc-*N*-methyl-4-iodophenyl alanine (both the *L*- and *D*-amino acids were used to explore the stereochemical preference at C-2),

followed by Suzuki coupling with various arylboronic acids $Ar^2B(OH)_2$ afforded the biaryl intermediate **5**. Removal of the Boc-protecting group with TFA followed by cyclization-release using triethylamine provided the des-amide biaryl diketopiperazine **6** (library A).

Next, as shown in Scheme 2, the synthesis of parallel library B (**10**) began with 2-nitrobenzene-sulfonyl-amino acid ester **7** that was prepared from esterification of TentaGelTM-S-OH resin with an Fmoc-amino acid (both the *L*- and *D*-amino acids were used to explore the stereochemical preference at C-5) using 2,6-dichlorobenzoyl chloride and pyridine followed by Fmoc deprotection using piperidine and subsequent re-protection with 2-nitrobenzene-sulfonyl chloride. Treatment of **7** with various primary alcohols R^1OH under Mitsunobu conditions (based on Liskamp's solid-phase adaptation⁴ of the Fukuyama secondary amine synthesis method⁵) provided **8**. Removal of the 2-nitrobenzene-sulfonyl group using PhSH and DBU in DMF followed by acylation with Boc-*N*-methyl-4-iodophenyl alanine (both the *L*- and *D*-amino acids were used to explore the stereochemical preference at C-2) and subsequent Suzuki coupling with arylboronic acids $Ar^2B(OH)_2$ under standard conditions afforded biaryl intermediate **9**. Deprotection of Boc and cyclization-release afforded the C-5 substituted biaryl diketopiperazine **10** (library B).

Finally, as also shown in Scheme 2, the synthesis of parallel library C began with biaryl intermediate **9** when $Ar^2 = 3$ -formylphenyl (aldehyde intermediate). Thus, reductive amination of the aldehyde intermediate with various R^4NH_2 followed by acylation with diverse R^5 derivatizing agents (acid chlorides, chloroformates, and isocyanates) and subsequent Boc-deprotection and cyclization-release produced the *meta*-substituted biaryl diketopiperazine **11** (library C).

Taken together, three parallel optimization libraries with over 300 compounds were prepared and each compound (~ 5 mg) was purified by HPLC prior to the CHO-hFSHR-luciferase assay.⁶ The assay results for



Scheme 2. Synthesis of parallel libraries B and C: (a) R^1OH , Ph_3P , DIAD, NMP, $-15^\circ C$ to rt; (b) PhSH, DBU, DMF; (c) Boc-*L*- or *D*-*N*-Me-4-iodophenylalanine, HATU, DIEA, DMF; (d) $Ar^2B(OH)_2$, $Pd(PPh_3)_4$, K_2CO_3 , DMF, $65^\circ C$; (e) TFA, CH_2Cl_2 ; (f) Et_3N , CH_2Cl_2 ; (g) R^4NH_2 , $NaBH(OAc)_3$, $ClCH_2CH_2Cl$; (h), R^5 derivatization (forming amides, carbamates, ureas).

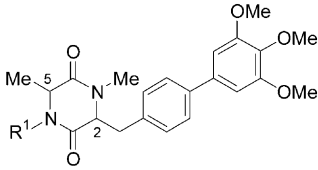
Table 1. EC_{50} values for select library A compounds in the CHO-hFSHR-luciferase assay

Compd	R^1	Ar^2	C-2	EC_{50} (nM)
6a	<i>n</i> -C ₉ H ₁₉	3,4,5-(MeO) ₃ C ₆ H ₂	<i>S</i>	900
6b	<i>n</i> -C ₈ H ₁₇	3,4,5-(MeO) ₃ C ₆ H ₂	<i>S</i>	400
6c	<i>n</i> -C ₇ H ₁₅	3,4,5-(MeO) ₃ C ₆ H ₂	<i>S</i>	2300
6d	<i>n</i> -C ₆ H ₁₃	3,4,5-(MeO) ₃ C ₆ H ₂	<i>S</i>	3000
6e	<i>n</i> -C ₅ H ₁₁	3,4,5-(MeO) ₃ C ₆ H ₂	<i>S</i>	9000
6f	<i>n</i> -C ₈ H ₁₇	3,4-(MeO) ₂ C ₆ H ₃	<i>S</i>	3300
6g	<i>n</i> -C ₈ H ₁₇	3,5-(MeO) ₂ C ₆ H ₃	<i>S</i>	5000
6h	<i>n</i> -C ₈ H ₁₇	3,5-Me ₂ -4-MeOC ₆ H ₂	<i>S</i>	1200
6i	<i>n</i> -C ₆ H ₁₃	3,4,5-(MeO) ₃ C ₆ H ₂	<i>R</i>	> 50,000

select compounds from libraries A, B, and C to highlight the key SAR are presented in Tables 1–3, respectively. (EC_{50} values are means of at least two experiments with standard deviations less than 20%.)

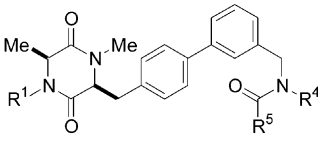
First, as is evident from Table 1, removal of the side chain amide bond is in fact beneficial. The des-amide compound **6a**, while having the same atom length for the *N*-4-substituent as in **1**, shows a 3-fold increase in potency relative to **1**. Shortening the *N*-4 side chain by one carbon atom gives rise to an additional 2-fold increase in potency (**6b**, EC_{50} = 400 nM). However, further truncation in chain length causes a gradual decrease in potency (**6c–e**). In addition, branched or cyclic alkyls at *N*-4 all result in inactive compounds (EC_{50} > 50 μ M, data not shown). Thus, the octyl group appears to be the optimal *N*-4-substituent. Furthermore, replacements of the 3,4,5-trimethoxyphenyl group with other aryls—3,4-dimethoxyphenyl (**6f**), 3,5-dimethylphenyl (**6g**), or 3,5-dimethyl-4-methoxy-phenyl (**6h**), for example—all result in less potent compounds (compare **6f–h** with **6b**). Also, similar to that in the original amide series,¹ the (*S*)-enantiomers in the des-amide series are consistently more potent than the (*R*)-enantiomers (as one example, compare **6d** with **i**).

Table 2. EC_{50} values for select library B compounds in the CHO-hFSHR-luciferase assay



Compd	C-5	C-2	R ¹	EC_{50} (nM)
10a	S	S	<i>n</i> -C ₆ H ₁₃	100
10b	S	S	<i>n</i> -C ₇ H ₁₅	25
10c	S	S	<i>n</i> -C ₈ H ₁₇	13
10d	R	S	<i>n</i> -C ₈ H ₁₇	120
10e	S	R	<i>n</i> -C ₈ H ₁₇	68
10f	R	R	<i>n</i> -C ₈ H ₁₇	1600

Table 3. EC_{50} values for select library C compounds in the CHO-hFSHR-luciferase assay



Compd	R ¹	R ⁴	R ⁵	EC_{50} (nM)
11a	<i>n</i> -C ₆ H ₁₃	Bu	Me	150
11b	<i>n</i> -C ₈ H ₁₇	Bu	Me	14
11c	<i>n</i> -C ₆ H ₁₃	Bu	OMe	8.9
11d	<i>n</i> -C ₈ H ₁₇	Bu	OMe	2.7
11e	<i>n</i> -C ₆ H ₁₃	Bu	NHMe	7.2
11f	<i>n</i> -C ₈ H ₁₇	Bu	NHMe	1.2
11g	<i>n</i> -C ₈ H ₁₇	MeOC ₂ H ₅	Me	9.7
11h	<i>n</i> -C ₈ H ₁₇	MeOC ₂ H ₅	OMe	1.8
11i	<i>n</i> -C ₈ H ₁₇	MeOC ₂ H ₅	NHMe	3.6

Second, as can be seen in Table 2, incorporation of a single (*S*)-methyl group at the C-5 position of the diketopiperazine ring results in a remarkable 30–100-fold increase in potency (compare **10a–c**⁷ with **6d–b**). However, compounds bearing bigger C-5-substituents—(*S*)-Et, Pr, or Bu—are all less potent than **10c** (data not shown), suggesting that the (*S*)-Me is the optimal substituent. In addition, the (2*S*,5*S*)-*cis*-stereochemistry affords the most active compound (**10c**, EC_{50} = 13 nM), relative to which, the corresponding (2*S*,5*R*)-*trans*-isomer (**10d**) has a 9-fold potency drop (EC_{50} = 120 nM); the (2*R*,5*S*)-*trans*-isomer (**10e**) a 5-fold potency drop (EC_{50} = 68 nM); and the (2*R*,5*R*)-*cis*-isomer (**10f**) a greater than 100-fold potency drop (EC_{50} = 1600 nM).

Finally, as shown in Table 3, similar to the trimethoxy series, the *meta*-aminomethyl series also shows a substantial potency enhancement with the incorporation of a single (5*S*)-Me on the diketopiperazine ring (**11a–i**, EC_{50} < 150 nM) relative to the C-5 unsubstituted compounds (EC_{50} > 1000 nM, data not shown). In addition, the *N*-4-octyl compounds (**11b,d,f**) all exhibit higher potency levels (by 3–10-fold) than the corresponding *N*-4-hexyl compounds (**11a,c,e**). Furthermore, the R⁴ butyl side chain can be replaced with other groups such as a methoxyethyl to provide equipotent compounds (**11g–i** versus **11b**, **11d**, **11f**). Finally, ureas and carbamates are generally more potent than their corresponding amides. For example, **11e** (urea) and **11c** (carbamate) are both >10-fold more potent than **11a** (amide).

Some of the most potent compounds were further evaluated in a CHO-hFSHR cell-based cAMP accumulation assay⁶ and the data is presented in Table 4. As can be seen, both the trimethoxy series (**10b–c**) and the *meta*-substituted series (**11b–f**) display agonistic activity in the cAMP accumulation assay, generating efficacies between 47% (**10b**) and 85% (**11e**) relative to FSH (100%).

In summary, we have described the optimization of low micromolar compounds **1** and **2** into potent low nanomolar FSH receptor agonists such as **10c** and **11f** through the design, synthesis, and evaluation of three parallel libraries with over 300 compounds. This work has demonstrated that parallel synthesis is a valuable tool for lead optimization in drug discovery. Further study of these compounds will be reported in due course.

Table 4. EC_{50} and efficacy values for compounds **10b–c** and **11b–f** in the CHO-hFSHR cell-based cAMP accumulation assay

Compd	EC_{50} (nM)	Efficacy (%)
10b	32	54
10c	32	47
11b	35	57
11c	48	59
11d	31	49
11e	9.5	85
11f	7.9	72

References and notes

1. Guo, T.; Adang, A. E. P.; Dolle, R. E.; Dong, G.; Fitzpatrick, D.; Geng, P.; Ho, K.-K.; Kultgen, S. G.; Liu, R.; McDonald, E.; McGuinness, B. F.; Saionz, K. W.; Valenzano, K. J.; van Straten, N. C. R.; Xie, D.; Webb, M. L. *Bioorg. Med. Chem. Lett.* **2004**, *14*, preceding paper in this issue. doi:10.1016/j.bmcl.2004.01.042.
2. Scott, B. O.; Siegmund, A. C.; Marlowe, C. K.; Pei, Y.; Spear, K. *Mol. Diversity* **1995**, *1*, 125.
3. Szardenings, A. K.; Burkoth, T. S.; Lu, H. H.; Tien, D. W.; Campbell, D. A. *Tetrahedron* **1997**, *53*, 6573.
4. Reichwein, J. F.; Liskamp, R. M. J. *Tetrahedron Lett.* **1998**, *39*, 1243.
5. Fukuyama, T.; Jow, C.-K.; Cheung, M. *Tetrahedron Lett.* **1995**, *36*, 6373.
6. (a) Brasier, A.; Tate, J.; Habener, J. *BioTechniques* **1989**, *7*, 1116. (b) Benzakour, O.; Kanthou, C.; Dennehy, U.; Al Haw, A.; Berg, L.-P.; Kakkar, V. V.; Cooper, D. N. *Biochem. J.* **1995**, *309*, 385.
7. Characteristic analytical data for **10c**: ^1H NMR (300 MHz, CDCl_3) δ 7.47 (d, 2H, $J=8.3$ Hz), 7.14 (d, 2H, $J=8.3$ Hz), 6.97 (s, 2H), 4.20 (t, 1H, $J=4.4$ Hz), 3.89 (s, 6H), 3.86 (s, 3H), 3.75 (q, 1H, $J=7.0$ Hz), 3.56 (m, 1H), 3.34 (dd, 1H, $J=4.2$ Hz, 13.8 Hz), 3.16 (dd, 1H, $J=4.2$ Hz, 13.8 Hz), 3.02 (s, 3H), 2.85 (m, 1H), 1.57 (m, 1H), 1.41 (m, 1H), 1.23 (m, 10H), 0.84 (t, 3H, $J=6.6$ Hz), 0.57 (d, 3H, $J=7.0$ Hz); MS (ESI): m/z 511.3 (MH^+).